Technical Note: A simple method for air–sea gas exchange measurements in mesocosms and its application in carbon budgeting

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Abstract. Mesocosms as large experimental units provide the opportunity to perform elemental mass balance calculations, e.g. to derive net biological turnover rates. However, the system is in most cases not closed at the water surface and gases exchange with the atmosphere. Previous attempts to budget carbon pools in mesocosms relied on educated guesses concerning the exchange of CO2 with the atmosphere. Here, we present a simple method for precise determination of air–sea gas exchange in mesocosms using N2O as a deliberate tracer. Beside the application for carbon budgeting, transfer velocities can be used to calculate exchange rates of any gas of known concentration, e.g. to calculate aquatic production rates of climate relevant trace gases. Using an arctic KOSMOS (Kiel Off Shore Mesocosms for future Ocean Simulation) experiment as an exemplary dataset, it is shown that the presented method improves accuracy of carbon budget estimates substantially. Methodology of manipulation, measurement, data processing and conversion to CO2 fluxes are explained. A theoretical discussion of prerequisites for precise gas exchange measurements provides a guideline for the applicability of the method under various experimental conditions.

1 Introduction

Pelagic mesocosms represent large volume (mostly between one and fifty m3) experimental enclosures used to gather data on natural plankton communities (Petersen et al., 2003). Generally open towards the atmosphere, mesocosms, however allowing for air–sea gas exchange, make it difficult to calculate production or consumption of CO2 and other volatile compounds inside an experimental unit. Climate relevant trace gases and other volatile carbon compounds produced in marine environments are increasingly investigated for their potential climate feedbacks and have been measured in previous mesocosm experiments (Sinha et al., 2007; Archer et al., 2012; Hopkins et al., 2012). Observed concentrations in a mesocosm are a product of water-column reactions and losses or gains from the atmosphere. Precise knowledge of air–sea gas exchange rates can be used to calculate net production rates of these compounds in the water column, which can be compared between various experiments. Aquatic production rates in concert with data on biological community composition and physiological state would help to understand observed open-ocean distributions.

Not only in the context of global change, biological CO2 fixation and consequent carbon export by sinking particles is of special interest to biogeochemical experimentalists. Most mesocosm studies currently focus on investigating ecological interactions applying standard oceanographic methods on subsamples of the enclosed water. In principal, mesocosm experiments also provide the opportunity to compare biogeochemical element fluxes such as air–sea gas exchange and export to water-column production. With production rates, as usually measured in side experiments (e.g. O2 production or 14C incorporation), uncertainties arise from sample transfer into bottle incubations and from extrapolating back from incubation conditions to temperature and light gradients present in mesocosms. In situ primary production measurements using the whole enclosure as experimental vessel have to be elaborated, in order to produce estimates comparable to total mesocosm fluxes like sedimentation of organic matter. Calculating carbon fluxes from water-column pools of inorganic and organic carbon quantitatively related to air–sea fluxes and export rates could largely improve the
understanding of the system (Czerny et al., 2012a). To directly estimate cumulative net community production (NCP), changes in total dissolved inorganic carbon (CT) have to be corrected for CO$_2$ air–sea gas exchange and eventually for calcification and evaporation. In previous mesocosm experiments in a Norwegian fjord (Delille et al., 2005) and indoors (Wohlers et al., 2009; Taucher et al., 2012), net community production (NCP), calculated on the basis of measured changes in CT, were presented. To calculate air–sea gas exchange, Delille et al. (2005) used a parameterisation for wind dependent boundary layer thickness achieved from experimental data compiled by Smith (1985). Wind speed, the crucial input parameter, was set to zero, because the mesocosms were closed to the atmosphere and moored in a sheltered surrounding. Whereas most parameterisations result in zero gas exchange at zero wind speed (Wanninkhof, 1992), laboratory derived wind dependent parameterisations by Smith et al. (1985) resulted in positive exchange at zero wind speed. Under calm conditions, gas exchange is low, but not zero; it is governed by other energy inputs than wind, e.g. thermal convection due to evaporation and temperature changes (Liss, 1973; Wanninkhof et al., 2009). Although direct wind forcing might be negligible in most mesocosms, the general assumption that overall energy input is comparable to the conditions in the experimental tanks used by Smith et al. (1985), however, is not justifiable. Surface turbulence in many mesocosm experiments is unlikely to be very low. Active mixing systems, wave movement of the surrounding water, thermal mixing or the deployment of sampling gear might create turbulence within the enclosures, comparable to quite windy conditions. Taucher et al. (2012), for example, found wind speeds of more than 6 m s$^{-1}$ to be necessary for balancing the carbon budget in a Kiel indoor mesocosm experiment, applying the Smith et al. (1985) calculation. Parameterisations for wind speed dependent gas exchange over the ocean are obviously not suitable for calculating mesocosm air–sea gas exchange. Other than open-ocean gas exchange measurements, direct measurement of exchange velocities in an enclosed water volume can be relatively easily done. Here, we present a simple method for direct measurements of air–sea gas exchange rates in mesocosm experiments using N$_2$O as a tracer. N$_2$O is a perfect choice as a gas tracer in this application, due to its well known atmospheric concentration, relatively simple detection and structural similarity to CO$_2$. Although N$_2$O is not an inert gas, conditions favouring its biological production are unlikely to occur in pelagic mesocosms. Possible bias by biological activity is assessed by parallel measurement of natural variations in N$_2$O and will be discussed later in the manuscript. The conversion of measured N$_2$O exchange rates to those of CO$_2$ and other gases is explained. We are providing a detailed description of the method and calculations including a discussion of prerequisites to achieve high quality data.

The measurement protocol and results are explained using a KOSMOS (Kiel Off Shore Mesocosm for Ocean Simula-

tion, Fig. 1) experiment on ocean acidification in the Arctic as a model. Applicability of the method in the Kiel indoor mesocosm facility is further explained and discussed.

2 Methods

2.1 Setup of the Svalbard 2010 ocean acidification experiment

Nine 15 m deep KOSMOS mesocosms, each with a diameter of 2m were moored end of May 2010 in the Kongsfjorden, Svalbard. Seven different CO$_2$ treatment concentrations were achieved through addition of CO$_2$ saturated seawater. While the ambient (\( \sim 180 \mu \text{atm} p\text{CO}_2 \)) control treatment was replicated twice, the seven enriched mesocosms followed a gradient up to \( \sim 1420 \mu \text{atm} p\text{CO}_2 \). Development of the enclosed natural plankton community was followed for 30 days after CO$_2$ manipulation, including addition of inorganic nutrients on day 13. For more details see Riebesell et al. (2012) and Schulz et al. (2013).

2.2 N$_2$O addition

One litre of saturated N$_2$O solution (N$_2$O medical, Air Liquide, purity > 98 \%) was prepared via bubbling of seawater for two days in a narrow measurement cylinder covered with Parafilm®. Amounts of the solution to be added to the mesocosms were calculated using solubility constants by Weiss and Price (1980) with respect to in situ salinities (S) and temperatures (T). The targeted concentrations of N$_2$O should be adapted to the setup in order to achieve mesocosm to air fluxes, which can be measured at good precision over reasonable time scales. Here, seawater tracer concentrations were chosen in accordance to the highest certified reference material for N$_2$O analyses available in our lab (\( \sim 55 \text{nmol kg}^{-1} \)).

Assuming a background concentration of 13 nmol kg$^{-1}$, 40 nmol kg$^{-1}$ of medical grade N$_2$O was added. Based on experience, a surplus of approximately 20 % was added to the mesocosms to account for losses unavoidable during handling of the solution.

Addition of the solution to the mesocosms (about 1–2 mL m$^{-3}$) can be calculated according to the formulation:

\[
V_{ad} = \frac{V_w \cdot ad}{K_{TS} \cdot p},
\]

(1)

where \( V_{ad} \) is the volume of N$_2$O stock solution added (L), \( V_w \) the volume of the mesocosm (L), ad the desired addition (mol L$^{-1}$), and \( K_{TS} \) is the solubility constant by Weiss and Price (1980) for S and T of the N$_2$O stock solution (mol L$^{-1}$ atm$^{-1}$) prepared at a pressure, \( p \) (atm), of one atmosphere.

A syringe with a large inlet diameter was used to transfer the stock solution carefully. Filling of the syringe was done
slowly as vacuum increases undesired outgassing of N$_2$O. The stock solution was first diluted with filtered seawater in 25 L carboys, which were filled almost to the rim. The content of the carboys was homogeneously distributed to the mesocosms by using the pumped injection device “Spider” (Riebesell et al., 2012).

2.3 Sampling

Three of the nine mesocosms were sampled every second day using integrating water samplers (IWS, Hydrobios). Equal amounts of sample were sucked into the sampling bottle at each depth between 0 and 12 m, electronically controlled via hydrostatic pressure sensors. These integrated water samples represent inventories of the 15 m deep water column. Triplicate samples were drawn directly from the sampler. The water was filled bubble free into 50 mL headspace vials via a hose reaching to the bottom of the vial. The vial volume was allowed to overflow about four times before closing. Vials were closed with butyl rubber plugs (N 20, Machery and Nagel), crimp sealed and stored at room temperature after addition of 50 µL of saturated mercury chloride solution.

2.4 Measurement procedures

Measurement of aquatic N$_2$O concentrations was performed via gas chromatography (GC) with electron capture detection (Hewlett Packard 5890 II), using a headspace static equilibration procedure as described by Walter et al. (2006, precision $\sim \pm 1.8\%$). The GC was equipped with a 6’/1/8’’ stainless steel column packed with a 5 Å molecular sieve (W. R. Grace & CO) and operated at a constant oven temperature of 190 °C. A 95/5 argon-methane mixture (5.0, Air Liquide) was used as carrier gas. 10 mL of helium (5.0, Air Liquide) headspace was added to the sample vials and later injected into the sample loop of the GC after equilibration was achieved by manual shaking and storage of the vials for at least 10 h at a temperature of 21 °C. Certified gas mixtures of N$_2$O in artificial air (Deuste Steininger GmbH) with mixing ratios of 87.2 ± 0.2, 318 ± 0.2 and 1008 ± 0.2 ppb as well as 1 : 1 dilutions with helium were used to construct calibration curves with a minimum of three data points close to sample concentrations. Headspace to water phase ratios in the vials was determined gravimetrically.

Total dissolved inorganic carbon (CT) was determined via coulometric titration using a SOMMA system and total alkalinity (TAlk) via potentiometric titration (Dickson, 1981) (standard error of both methods $\sim \pm 1 \mu$mol kg$^{-1}$). CO$_2$ concentrations, partial pressures and pH (total scale) were calculated from CT and TAlk measurements with the program CO2SYS by Lewis and Wallace (1995). For more details on carbonate chemistry see Bellerby et al. (2012).

Determination of salinity and temperature in the mesocosms was performed with a data logger-equipped hand held multisensory CTD 60M (Sea and Sun Technology). Volume
of the mesocosms was determined with the same instrument using sodium chloride additions of \( \sim 0.2 \text{ g kg}^{-1} \) as a tracer (Czerny et al., 2012b).

Wind velocity and direction measured at 10 m height on-shore, about one mile from the mooring site, were provided by the staff of the AWI-PEV Station in Ny Alesund.

Atmospheric measurements of \( \text{N}_2\text{O} \) and \( \text{CO}_2 \) were measured on close by Zeppelin Mountain (\( \sim 4.5 \text{ km} \) from the experimental side) and provided to us by the NOAA Carbon Cycles Gases Group in Boulder, CO, USA and ITM in Stockholm University, Sweden, for \( \text{N}_2\text{O} \) and \( \text{CO}_2 \), respectively.

### 3 Results and discussion

Concentrations of \( \text{N}_2\text{O} \) added on day 4 decreased in the enriched mesocosms from initially measured \( \sim 50 \text{ nmol kg}^{-1} \) on day 6 to \( \sim 30 \text{ nmol kg}^{-1} \) on day 28 (Fig. 2). Concentrations measured in the fjord close to the mesocosms were slightly oversaturated compared to atmospheric equilibrium values, calculated for in situ seawater \( T, S \) and atmospheric mixing ratios measured close by on Zeppelin Mountain. Despite variable wind conditions, the concentration decrease inside the mesocosms could be fitted \((R^2 = 0.96)\) using a standard diffusion relationship:

\[
C_{\text{N}_2\text{O}} = 60.556 \cdot e^{-0.0241 \cdot d} \tag{2}
\]

where the concentration of \( \text{N}_2\text{O} \) \((C_{\text{N}_2\text{O}})\) is described as an exponential function of the sampling day \((d)\).

#### 3.1 Calculation of \( \text{CO}_2 \) fluxes from changes in \( \text{N}_2\text{O} \) concentrations

Daily \( \text{N}_2\text{O} \) fluxes were calculated from the fitted \( \text{N}_2\text{O} \) concentration decrease over time and converted to volumetric units. Changes in the \( \text{N}_2\text{O} \) inventory, derived using the determined volume of the mesocosms (method described in Czerny et al., 2012b) were used to calculate fluxes in \( \text{µmol cm}^{-2} \text{ h}^{-1} \) \((F_{\text{N}_2\text{O}})\) across the water surface according to

\[
F_{\text{N}_2\text{O}} = \frac{I_{w1} - I_{w2}}{A \cdot \Delta t} \tag{3}
\]

where \( I_{w1} \) is the fitted bulk water \( \text{N}_2\text{O} \) inventory in \( \text{µmol per mesocosm} \) on \( t_1 \) and \( I_{w2} \) on \( t_2 \) with \( \Delta t \) as the time interval between \( t_1 \) and \( t_2 \) in \( \text{h} \), while \( A \) is the nominal surface area of the mesocosm in \( \text{cm}^2 \). A \( \text{N}_2\text{O} \) transfer velocity \((k_{\text{N}_2\text{O}})\) in \( \text{cm h}^{-1} \) is then calculated by dividing \( F_{\text{N}_2\text{O}} \) by the concentration gradient according to Eq. (4):

\[
k_{\text{N}_2\text{O}} = \frac{F_{\text{N}_2\text{O}}}{(C_{\text{N}_2\text{O} \text{w}} - C_{\text{N}_2\text{O} \text{aw}})} \tag{4}
\]

where \( C_{\text{N}_2\text{O} \text{w}} \) is the fitted bulk water \( \text{N}_2\text{O} \) concentration \((\text{µmol cm}^{-3})\) at the point in time and \( C_{\text{N}_2\text{O} \text{aw}} \) the calculated (Weiss and Price, 1980) equilibrium concentration of \( \text{N}_2\text{O} \) with the atmosphere at prevailing bulk water \( T \) and \( S \). Using bulk water concentrations to derive a surface diffusion gradient perfect mixing of the mesocosm appears to be an absolute requirement. Referring from \( \text{N}_2\text{O} \) to inert gasses with air–sea gas exchange being the only exchange process, it is irrelevant whether exchange is limited by mixing processes close to the air–sea interface or within the water column. Yet, if a permanent stratification is formed inside the mesocosm, the decrease of \( \text{N}_2\text{O} \) bulk water concentration cannot be used to calculate mesocosm–atmosphere \( \text{CO}_2 \) exchange. Processes modulating the concentrations of biologically active compounds such as \( \text{CO}_2 \) are usually variable along the light gradient. Therefore, due to shallow primary production, considerable differences in the surface gradient of \( \text{CO}_2 \) might emerge compared to \( \text{N}_2\text{O} \) surface gradients that are governed by diapycnal mixing. For stratified mesocosms, gas exchange calculations require the integration of information about vertical distribution of tracer and gases of interest. Here, \( \text{N}_2\text{O} \) and \( \text{CO}_2 \) inventories have to be determined by integrated water samples independently from surface gradients determined from discrete surface water samples. Regardless of the specific sampling strategy applied, it is imperative to use the same protocol for the tracer as for the gases of interest.

Due to convection caused by slight temperature changes in the surrounding water (Fig. 5) and an evaporation induced salinity increase (Schulz et al., 2013), the mesocosms in the Svalbard KOSMOS study could be considered to be homogeneous on time scales relevant for air–sea gas exchange. \( \text{N}_2\text{O} \) as well as \( \text{CO}_2 \) surface concentrations are therefore adequately represented by bulk water measurements.

\( k_{\text{N}_2\text{O}} \) can be translated into a transfer velocity for any other gas using its Schmidt numbers to correct for gas specific
The Schmidt number for N\(_2\)O (\(S_{N_2O}\)) published by Rhee (2000), and the Schmidt number for CO\(_2\) (\(S_{CO_2}\)) derived from diffusion coefficients published by Jähne et al. (1987) were used. Using this pair of Schmidt numbers, \(k_{CO_2}\) is generally less than 1 % smaller than \(k_{N_2O}\), similarly to when both Schmidt numbers are derived from coefficients published by Jähne et al. (1987). If both coefficients are taken from Wilke and Chang (1955) the difference is ∼ 10 % and can become larger than 25 % if \(S_{CO_2}\) from Wilke and Chang (1955) is paired with \((S_{N_2O})\) derived from coefficients published by Broecker and Peng (1974). More recent wind and wave tank experiments have shown that a conversion of \(k_{N_2O}\) to \(k_{CO_2}\) is not necessary as gas exchange of the two gases is indistinguishable under conditions were chemical enhancement of CO\(_2\) exchange is not relevant (Degreif, 2006).

Fluxes for CO\(_2\) (\(F_{CO_2}\)) can then be calculated by multiplication of \(k_{CO_2}\) with the diffusion gradient between bulk water CO\(_2\) concentrations \((C_{CO_2w})\) and calculated equilibrium concentrations with the atmosphere \((C_{CO_2a})\) as

\[
F_{CO_2} = k_{CO_2} \cdot (C_{CO_2w} - C_{CO_2a}).
\]

Daily CO\(_2\) fluxes were calculated for the nine mesocosms within the Svalbard ocean acidification experiment (Fig. 3). In the first days after CO\(_2\) addition was completed (day 4), maximum efflux of ∼2 µmol CO\(_2\) per kg seawater and day could be observed in the highest CO\(_2\) treatment (∼1400 µatm) at a CO\(_2\) gradient of ∼980 µatm. In the following two weeks, the CO\(_2\) gradient was diminished by outgassing CO\(_2\) in concert with biological uptake, so that fluxes on day 27 were considerably lower (gradient ∼450 µatm). Decrease of fluxes as a result of decreasing CO\(_2\) gradients was less pronounced in the more moderately oversaturated mesocosms due to a higher buffer capacity of the carbonate system. About 0.5 µmol kg\(^{-1}\) d\(^{-1}\) CO\(_2\) gassed into the water from the atmosphere in the non-manipulated control treatments (∼175 µatm). Here, biological uptake was roughly balanced by influx so that the gradient remained rather constant over time.

### 3.2 Chemical enhancement of CO\(_2\) air–sea gas exchange

Another correction has to be applied to derive accurate CO\(_2\) fluxes in calm environments like the KOSMOS mesocosms. As CO\(_2\) reacts with water, unlike N\(_2\)O, CO\(_2\) gas exchange might be chemically enhanced due to buffering of diffusive concentration change by equilibration reactions within the boundary layer. Other than inert gases, CO\(_2\) diffuses are not necessarily exchanged through the boundary layer, but can also be formed from bicarbonate close to the interface. This applies only at low wind speeds and not when mixing is considerably faster because CO\(_2\) hydration kinetics are slow. Thus, chemical enhancement is thought to be rather insignificant under turbulent conditions relevant for open-ocean CO\(_2\) exchange (e.g. when \(k > 5\) cm h\(^{-1}\)), but applies to the conditions found inside the mesocosms (\(k \sim 1.8–2.5\) cm h\(^{-1}\)) (Wanninkhof and Knox, 1996). Moreover, the state of the carbonate system determines the extent of chemical enhancement, being negligible at pH < 6 and substantial at pH > 8. In the Svalbard ocean acidification experiment, the treatment pH\(_{tot}\) (total scale) ranged from 7.5 to 8.3 (Bellerby et al., 2012), therefore chemical enhancement created a pH effect on carbon flows that must be considered. To correct for this, theoretical parameterisations by Hoover and Berkshire (1969) were chosen, as currently no empirical parameterisations exist sufficiently describing the process in natural seawater (Wanninkhof and Knox, 1996). The enhancement factor \(\alpha\), the ratio between chemical enhanced flux and not enhanced flux can be calculated using Eq. (7):

\[
\alpha = \frac{\tau}{[\tau - 1 + \tanh{(Qz)}] / (Qz)}.
\]

Here, dimensionless \(\tau = 1 + [H^+]^2 (K^+_w K^+_\text{H} + K^+_{\text{H}^+})^{-1}\), with \(K^+_w\) and \(K^+_{\text{H}^+}\) being the first and second stoichiometric equilibrium constants for carbonic acid and \([H^+]\) the proton concentration. \(Q = (rD)^{0.5}\) in cm\(^{-1}\), where \(D\) is the diffusion coefficient for CO\(_2\) by Jähne et al. (1987) and \(r\) describes the hydration of CO\(_2\) either directly or via true carbonic acid. \(r\) in the unit s\(^{-1}\) can be calculated using Eq. (8):

\[
r = K_{CO_2} + K_{OH^-} K^+_w [H^+]^{-1}
\]

with \(K_{CO_2}\) being the CO\(_2\) hydration rate constant (s\(^{-1}\)), \(K_{OH^-}\) is the CO\(_2\) hydroxylation rate constant (L mol\(^{-1}\) s\(^{-1}\)).
from Johnson (1982) and $K_w^+$ is the equilibrium constant for water. The boundary layer thickness $z$ (cm) can be calculated from determined transfer velocity ($z = Dk_{CO_2}^{-1}$). All constants used here can be found in Zeebe and Wolf-Gladrow (2001). Using the Hoover and Berkshire (1969) model, input conditions similar to our experimental conditions in Svalbard ($T = 5^\circ$C, $S = 35$, $z = 0.002$ cm, $pH_{tot} = 8.2$) result in enhancement of about 8% ($\alpha = 1.082$). For the same conditions, but at a temperature of 25$^\circ$C, CO$_2$ gas exchange would be enhanced by about 48% ($\alpha = 1.479$).

Chemical enhancement factors using more complex models published by Quinn and Otto (1971), Emerson (1975), Smith (1985), and Keller (1994) give very similar results to the Hoover and Berkshire (1969) model (Wanninkhof and Knox, 1996). Experimental data from tank experiments reproduce calculated chemical enhancement relatively well (i.e. Hoover and Berkshire, 1969; Liss, 1973; Wanninkhof and Knox, 1996; Degreif, 2006). The simple pH dependent fit derived from enhancement experiments in natural Baltic seawater published by Kuss and Schneider (2004) is not recommended for use, as influences of $T$, $S$ and $z$ are not considered.

The relevance of chemical enhancement for open-ocean CO$_2$ exchange is controversial as the calculation of $k$ from wind speed over the ocean itself still bears considerable uncertainty. As $k$ in our experiments is measured directly, comparability to experimental results is quite straightforward.

Due to low temperatures during the Svalbard experiment, chemical enhancement of $\sim 3$ to 7% is very low (Fig. 7). The influence of about three degree warming during the experiment in June 2010 is overall larger than the calculated difference arising from pH treatments (Fig. 4). Wrong pH-dependent chemical enhancement could produce artificial treatment effects in the carbon budget estimates especially in warm water ocean acidification studies. NCP estimates within this experiment by Silyakova et al. (2012) and Czerny et al. (2012a) at arctic temperatures are relatively unaffected by enhancement of this magnitude and possible uncertainties therein.

Evidence for a strong increase in chemical enhancement due to enzymatic catalysis by free carbonic anhydrases as suggested by Berger and Libby (1969) was not found in later experiments (Goldman and Denny, 1983; Williams, 1983), but it might be interesting to reconsider this question in future mesocosm experiments.

The lack of empirical data coverage on chemical enhancement parameterisations in seawater poses the major quantitative uncertainty for NCP estimates based on CO$_2$ air–sea gas exchange using the presented method. Especially in setups were temperatures are high, the proportion of CO$_2$ exchange relying on theoretical considerations is high compared to the directly measured flux.

### 3.3 The choice of N$_2$O as a gas exchange tracer and its biological stability

The N$_2$O molecule strongly resembles CO$_2$ in most physical properties; it has the same mass, nearly the same solubility and diffusivity. Other than CO$_2$, equilibrium reaction of N$_2$O with water lies strongly on the side of free N$_2$O so that air–sea gas exchange can be approximated as for inert gases. In laboratory experiments N$_2$O is a perfect tracer for CO$_2$ gas exchange. Because of the similarity of both gases a conversion of measured $k_{N_2O}$ to $k_{CO_2}$ is small and so are potential uncertainties. Wall effects relevant in laboratory experiments such as permeability or adhesion to plastic walls can be assumed to be comparable between similar molecules. In open waters, background concentrations of N$_2$O are slightly variable. Therefore, $^3$He and SF$_6$ were used as gas exchange tracers in open-ocean applications as they are highly inert, their natural background concentration and detection limit is very low so that measurement is possible also after considerable dilution. Despite many practical advantages of N$_2$O in the application in mesocosms its prominent role as a biologically produced climate relevant trace gas is putting the inertness of N$_2$O into question.

The natural source of oceanic background N$_2$O concentrations is biological production. N$_2$O is produced predominantly as a side product of nitrification, when ammonia is incompletely oxidised in the course of deep remineralisation at low oxygen concentrations. Yet, most parts of the ocean are near equilibrium with the atmosphere (mean global saturation 103%) (see Bange et al., 1996 and references herein), whereas significant N$_2$O oversaturation is predominantly found in tropical regions rather than in cold and temperate waters (Walter et al., 2006). Detectable nitrification in the euphotic zone was hypothesised to also be a source
of N\textsubscript{2}O (Dore and Karl, 1996; Santoro et al., 2010), but this was not yet directly observed. Physiological results (Goreau et al., 1980; Loescher et al., 2012) suggest possible N\textsubscript{2}O production by nitrification in fully oxygenated waters to be very low. However, even N\textsubscript{2}O production at relatively high surface layer nitrification rates, as found in upwelling regions (Rees et al., 2011), are orders of magnitudes too low to significantly bias the large fluxes caused by deliberate N\textsubscript{2}O addition. The only known pathway of biological N\textsubscript{2}O uptake as a reactive nitrogen species is by denitrifiers at anoxic conditions (\lless 10 \textmu mol O\textsubscript{2} kg\textsuperscript{-1}) (Zumft, 1997; Zamora et al., 2012). Conditions favouring this process are unlikely to form in mesocosms and would result in a loss of the tracer (as N\textsubscript{2}) but not into utilisation of N\textsubscript{2}O as a nitrogen source. Side-effects of N\textsubscript{2}O addition on biological activity in mesocosm experiments can therefore not be expected. Remineralisation of detritus at the bottom of the mesocosm could possibly be a source of N\textsubscript{2}O. Conditions allowing for extensive remineralisation of accumulated organics inside pelagic mesocosms should thus be avoided. It is further strongly recommended to measure background natural N\textsubscript{2}O concentrations preferably inside non-enriched experimental units, because N\textsubscript{2}O is not considered as an inert gas.

3.4 Lateral gas exchange by diffusion through mesocosm wall material

Gas fluxes through the mesocosm walls can be calculated if temperature dependent permeability coefficients of the foil material and gases of interest are known.

Fluxes through the wall (\(F\text{\textsubscript{wall}}\)) in mol d\textsuperscript{-1} can be derived using the equation:

\[
F\text{\textsubscript{wall}} = \frac{D_{\text{f}} \cdot \Delta p \cdot A \cdot t}{z_m}.
\]

Here \(D_{\text{f}}\) is the permeability coefficient at a given temperature in mol \cdot \mu m atm\textsuperscript{-1} m\textsuperscript{-2} d\textsuperscript{-1}, \(\Delta p\) is the partial pressure difference between inside and outside in atm, \(A\) is the submerged surface of the mesocosm walls in m\textsuperscript{2}, \(t\) is the duration in days and \(z_m\) is the thickness of the material in \mu m. Turbulence of the media in and outside the mesocosm is not relevant to this diffusion gradient as permeation of the materials is generally orders of magnitude slower than removal and advection of gases in the media.

Estimates of lateral gas exchange for the Svalbard experiment were calculated based on permeability coefficients published for Desmopan\textsuperscript{®} 385, (Bayer). Desmopan\textsuperscript{®} 385 is the raw material of our bag foil (Walopur\textsuperscript{®}, Eurepex Films). Direct measurements for Walopur\textsuperscript{®} are not available for CO\textsubscript{2} and N\textsubscript{2}O. Permeability for the specific temperatures was extrapolated.

For the experiments in the KOSMOS mesocosms, the fraction of the measured gas flux due to permeability of the bag material was maximal on the order of 1–2 %. Fluxes are low because of the relatively thick foil (0.5 to 1 mm) at comparatively low temperatures. In the perspective of CO\textsubscript{2} gas exchange estimates for carbon mass balance it is generally not interesting whether CO\textsubscript{2} exchanges through the walls or via the water surface. However, differences between N\textsubscript{2}O and CO\textsubscript{2} in the material specific permeability of the bags have the potential to cause systematic errors if exchange is largely through the foil and not with the atmosphere. Such differences seemed at first unlikely because of the general similarity of N\textsubscript{2}O and CO\textsubscript{2} in diffusivity and solubility, but permeability specifications for Desmopan\textsuperscript{®} 385 suggest a considerably higher permeability for N\textsubscript{2}O (27.1 and 51.7 mol \cdot \mu m atm\textsuperscript{-1} m\textsuperscript{-2} d\textsuperscript{-1} for CO\textsubscript{2} and N\textsubscript{2}O at 25 °C, respectively) (Bayer MaterialScience, TPU TechCentre). For the Svalbard experiment bias through lateral gas fluxes were not corrected, as the overall magnitude of these fluxes was negligible. The data basis in terms of permeability measurements would not have allowed for an exact correction of such bias. A set of permeability measurements at a relevant temperature range would improve gas exchange estimates especially at temperatures above 10 °C. If thin foil is used for mesocosms, a material with good gas barrier properties should be chosen and exact permeabilities should be known for the gases of interest.

When \(k_{\text{N2O}}\) is translated into transfer velocities of poorly water soluble gases, dissolution and adhesion of those gases in and on the plastic material could cause a lateral sink of these substances in addition to the permeability issue.

3.5 Sensibilities of the results towards uncertainties in measured variables

Sensitivities of the overall resulting CO\textsubscript{2} fluxes to uncertainties in the determination of the most important measured variables for the presented method were estimated (Table 1). Water temperatures are used on numerous occasions for the calculation of gas exchange rates, e.g. for the calculation of CO\textsubscript{2} from CT and TAlk, for Schmidt numbers, for solubility
Table 1. Sensitivities of the overall resulting CO$_2$ fluxes to uncertainties in the determination of the most important measured variables of the presented method. The effect of systematic N$_2$O underestimation was tested using an alternative fit including only upper end values from the Svalbard dataset. The influence of errors in CO$_2$ gradient determination is denoted for an intermediate gradient of 400 $\mu$atm.

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Uncertainty in parameter</th>
<th>Uncertainty in CO$_2$ fluxes</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sea surface temperature</td>
<td>±1 $^\circ$C</td>
<td>±3 %</td>
</tr>
<tr>
<td>Mesocosm volume</td>
<td>±1 %</td>
<td>±1 %</td>
</tr>
<tr>
<td>Mesocosm surface</td>
<td>±1 %</td>
<td>±1 %</td>
</tr>
<tr>
<td>Systematic error in N$_2$O measurement</td>
<td>Outliers due to losses during sampling</td>
<td>0.26 ± 0.29 %</td>
</tr>
<tr>
<td>Air–sea CO$_2$ gradient</td>
<td>±2 $\mu$mol in CT and TAlk</td>
<td>±5 %</td>
</tr>
</tbody>
</table>

and chemical enhancement. Errors in $k_{CO_2}$, in response to uncertainties in temperature, are mainly caused by the temperature dependence of N$_2$O solubility. Errors in resulting CO$_2$ fluxes are largely balanced by errors in CO$_2$ solubility calculated using identical temperatures. The remaining sensitivity of 3 % for 1 $^\circ$C appears to be relatively low in respect of usually very precise temperature measurements using current technology. It has to be kept in mind that gas exchange is a continuous process; therefore, measurement frequency should be adequate for sufficient description of relevant temperature changes during the entire duration of the experimental duration. At water temperatures above 10 $^\circ$C, chemical enhancement corrections become important so that precise temperature records gain additional relevance. Uncertainties in mesocosm volume or surface area translate directly into errors in calculated CO$_2$ fluxes (one to one %) when $k_{CO_2}$ measured in one mesocosm is applied to calculate gas exchange in a parallel mesocosm with different volume or surface area. A random uncertainty in N$_2$O determination of 1.8 % as denoted by Walter et al. (2006) would be averaged out due to the large number of fitted measurements. However, a hypothetical systematic underestimation of N$_2$O by including outliers possibly caused by N$_2$O losses from oversaturated samples would influence $k_{CO_2}$. The effect of systematic N$_2$O underestimation was tested using an alternative fit including only upper end values. Resulting CO$_2$ fluxes differ by 0.7 % in the beginning of the experiment when N$_2$O concentrations were high and roughly equal fluxes calculated including all N$_2$O measurements at the end of the experiment. Maximum uncertainties in CO$_2$ fluxes of 5 % due to errors in the determination of air–sea CO$_2$ gradients are calculated on the basis of a maximum uncertainty of ±$2 \mu$mol kg$^{-1}$ in CT as well as TAlk and an intermediate CO$_2$ gradient of ~400 $\mu$atm.

when gradients are large and vice versa. The effects of random errors in CO$_2$ determination on uncertainties in cumulative CO$_2$ mass flux over time are averaged out over time. A cumulative error of the applied constants cannot be given but it has to be highlighted that this is an additional source of uncertainty. Parameterisations of Schmidt numbers, solubility and rate constants, as well as diffusion coefficients cited in the text, were chosen to the best of our knowledge.

3.6 Processes driving gas exchange in mesocosms

The concentration of N$_2$O (C$_N$O) decreased quite steadily over the whole experimental period (Fig. 2). This indicates that N$_2$O fluxes were controlled by the diffusion gradient to the atmosphere. Variable external forcing by wind or waves as commonly observed in natural environments was of minor importance. Wind measurements at Bellevaja station at 10 m above sea level ($U_{10}$) reported velocities of up to 5 m s$^{-1}$ during the experiment (Fig. 6). The water surface of the mesocosms, however, is sheltered from direct wind shear by the two meter high plastic walls of the bag (Fig. 1; Riebesell et al., 2012).

Fetch, the distance wind could act on the water surface, was dependent on wind direction (maximum of ~10 nm in the Kongsfjorden). Waves that were able to propagate through the mesocosms (significant wave height ($H_{1/3}$) up to ~0.8 m) were only observed on the mooring site on three days when stronger winds were blowing along the fjord from southeast, the most exposed wind direction. Enhanced gas exchange during the days with waves could not be resolved by our measurements. However, CO$_2$ gas exchange inside the mesocosms was measured to be constantly about three times higher than calculated flux at zero wind as performed by Delille et al. (2005) (Fig. 7, stagnant film thickness calculated according to Smith, 1985, chemical enhancement according to Hoover and Berkshire, 1969).

Applying a quadratic wind dependent function (Wanninkhof, 1992) at constant wind speed of 3.15 m s$^{-1}$, resulting fluxes are very close to our empirical estimate over the whole period. Measured $U_{10}$ wind speeds at the experimental site were generally lower than this (mean 2.1 m s$^{-1}$), and accordingly calculated mean air–sea gas exchange was also lower outside in the fjord than inside the mesocosms. Compared to relevant open-ocean gas transfer, estimated mesocosm CO$_2$ transfer velocities between 1.9 to 2.5 cm h$^{-1}$ in the Svalbard experiment are low. They are within a gray zone of baseline gas exchange where buoyancy fluxes and chemical enhancement contribute largely to gas exchange so that purely wind depended parameterisations are not applicable (Wanninkhof et al., 2009). Additional factors can be argued to be driving gas exchange in mesocosms compared to open waters. Rinsing of the plastic walls when waves are propagating through the setup presumably leads to enhanced air–sea surface renewal compared to open water. Slight temperature changes in the surrounding water mass were immediately
heating or cooling the bags (Fig. 5), this probably caused considerably enhanced buoyancy fluxes that kept the experimental units relatively homogenous throughout the experiment. Last but not least, the extensive daily sampling with water samplers and probes contributed to gas exchange by active perturbation of the mesocosm surface.

3.7 Mesocosm proportions

Transfer velocities ($k$) in other mesocosm setups deployed in more sheltered surroundings, standing on land or inside climate controlled rooms might be lower or higher, depending on methodology used for sampling, temperature control, active mixing and gas specific permeability of the mesocosm material. Even more important than these influences on $k$, is the ratio between the mesocosm volume and its surface area ($A/V$), when exchange rates are normalised to units of water (kg$^{-1}$ or L$^{-1}$). In an exemplary 15 m deep KOSMOS mesocosm (Fig. 1), holding $\sim 45 \text{ m}^3$ of water, CO$_2$ gas exchange over $3.14 \text{ m}^2$ ($A/V = 0.07$) surface area is causing relatively moderate changes in aquatic concentrations despite large diffusion gradients (Fig. 3). Taking the example of the Kiel indoor mesocosm (Fig. 8a) of about $1.4 \text{ m}^3$ at $2 \text{ m}^2$ surface ($A/V = 1.4$), concentration change in response to the same gas exchange flux is 20 times faster. Additionally, air–sea gas exchange velocities are accelerated by continuous active mixing, necessary to keep plankton organisms in suspension (Fig. 8a). While after 20 days $\sim 50\%$ of the N$_2$O added was still present during the Svalbard study (Fig. 1), the same tracer concentration was virtually gone after five days in the shallow indoor mesocosm (Fig. 8b) in the uncovered configuration. Here, inorganic carbon uptake by phytoplankton can be rapidly compensated by ingassing of CO$_2$ from the atmosphere. Ocean acidification experiments in setups with $A/V$ similar to the Kiel indoor mesocosm would lose their treatment CO$_2$ within a few days. To maintain treatment levels in such shallow experiments, continuous measurement and control technology can be used (e.g. Widdicombe and Needham, 2007). Resulting controlled treatment levels are beneficial when physiological questions are investigated. However, CO$_2$ drawdown does not occur, and therefore DIC concentration change cannot be used to calculate NCP. Another option is to artificially decrease the surface area by covering the mesocosm with a low permeability transparent film. For comparison 50 nmol kg$^{-1}$ N$_2$O was added to two Kiel indoor mesocosms, one in an uncovered configuration and one covered with a transparent floating foil to reduce surface in contact with the atmosphere (Fig. 8a). Both mesocosms were stirred at the same speed; samples were drawn using a tube. The thin polyurethane foil mounted on a light frame and floating on the surface, efficiently minimised air–sea gas exchange (Fig. 8b). If covers are used, reducing the surface area to a minimum, it has to be considered that the remaining open surface should be equally large. As the working principle of this approach is to minimise surface area, it can be assumed to be very sensible to the size of the remaining interface (leaks). Therefore, air–sea gas exchange should be measured in all experimental units to check for reproducibility of results.
4 Conclusion and outlook

Direct measurement of N$_2$O air–sea gas exchange can be used to estimate accurate CO$_2$ fluxes in various mesocosm setups, whereas common wind dependent parameterisations for air–sea gas exchange cannot be adapted to mesocosm conditions and their application is therefore prone to systematic errors. Within the mesocosm low energy physical surrounding, N$_2$O gas exchange measurements allow for direct estimation of CO$_2$ fluxes with uncertainties far below open-ocean dual tracer measurements due to a known, constant water volume. Measured transfer velocities are within the range of zero wind open-ocean baseline velocities assumed to be dominated by chemical enhancement and buoyancy fluxes (Wanninkhof et al., 2009). The influence of sea surface microlayers of surface active organic molecules is discussed to be responsible for large discrepancies in gas exchange between productive coastal waters and open-ocean conditions (Frew, 1997; Kock et al., 2012). The decrease in open-ocean $k$ by 20–50 %, due to surfactants smoothening wind effects on the water surface (Tsai and Liu, 2003), can be expected to be of minor relevance to mesocosm gas exchange where wind stress is not the dominating energy input. The effect of these surfactants, possibly produced in high amounts during phytoplankton blooms in mesocosms, is difficult to include in theoretical calculations, but is inherently included in our direct measurements. Future mesocosm experiments combining the close observation of biological, chemical and physical processes might offer the opportunity to bring more light into origin and composition of organic surface microlayers.

The application of gas exchange measurements for calculation of NCP inside the mesocosm delivered satisfying results. Of the four community production estimates published for the Svalbard 2010 experiment, NCP calculated from changes in dissolved inorganic carbon corrected for air–sea gas exchange (Czerny et al., 2012a; Silyakova et al., 2012) seems to be quantitatively most plausible. Although overall quantity compares relatively well with results from oxygen and in situ $^{13}$C–primary production estimates (Tanaka et al., 2013; de Kluijver et al., 2013), comparability to $^{14}$C incubation data presented by Engel et al. (2012) is weak. Much higher $^{14}$C fixation rates can be plausibly explained by the shallow ($\sim$1 m) incubation depth, while oxygen incubations obviously experienced more intermediate light and temperature conditions at $\sim$4 m depth. $^{13}$C tracer incorporation measurements inside the mesocosm deliver results representative for the entire mesocosm but only for a period before organic matter approached saturation with the tracer (de Kluijver et al., 2013). Rates measured in side experiments are more useful to compare community production between the treatments rather than giving quantitative cumulative estimates for in situ carbon uptake (see discussion in Engel et al., 2012). As incubations were performed only at one depth, it is impossible to integrate these data over time and depth in respect of variable light and temperature gradients. Summing up incubation results to achieve cumulative estimates could lead to further error propagation, whereas NCP calculated from in situ inorganic carbon measurements is per se cumulative and propagation of single measurement errors cannot occur.

Further development of the N$_2$O tracer concept is focussed on using it not only to determine air–sea gas exchange in stratified mesocosms, but also to estimate diapycnal mixing between surface layer and deep water inside mesocosms. For this purpose, N$_2$O gradients developing over time will be correlated to high resolution profiles of oxygen, pH and salinity, measured with CTD sensors. Especially in temperate turbid waters, mesocosm NCP is mostly not restricted by mesocosm length but by light penetration. The photoautotrophic surface layer communicates to some extent with a more nutrient rich deep layer where heterotrophic processes dominate. Budgeting these more naturally structured mesocosms is not only an interesting challenge, but will also introduce new ecological aspects connected to upward and downward elemental fluxes into the biogeochemical interpretation of the mesocosm system.

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Fig. 8. (a) Schematic drawing of the Kiel indoor mesocosm. With stirrer, floating lid and sampling hose, below: vertical view on floating foil lid with enforcement frame. (b) Comparison of N$_2$O tracer outgassing in a Kiel indoor mesocosm, between a simple uncovered setup (black) and a setup using a floating foil lid reducing the water surface area available for air–sea gas exchange (blue).
Stockholm University, Sweden, for providing us with atmospheric N\textsubscript{2}O and CO\textsubscript{2} measurements, respectively, from close by Zeppelin Mountain. We gratefully acknowledge the logistical support of Greenpeace International for its assistance with the transport of the mesocosm facility from Kiel to Ny-Ålesund and back to Kiel. We also thank the captains and crews of MV ESPERANZA of Greenpeace and R/V Viking Explorer of the University Centre in Svalbard (UNIS) for assistance during mesocosm transport and during deployment and recovery in Kongsfjorden. We thank the staff of the French-German Arctic Research Base at Ny-Ålesund, in particular Marcus Schumacher, for on-site logistical support.

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References


