Simulating the global distribution of nitrogen isotopes in the ocean

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[1] We present a new nitrogen isotope model incorporated into the three-dimensional ocean component of a global Earth system climate model designed for millennial timescale simulations. The model includes prognostic tracers for the two stable nitrogen isotopes, 14N and 15N, in the nitrate (NO3−), phytoplankton, zooplankton, and detritus variables of the marine ecosystem model. The isotope effects of algal NO3− uptake, nitrogen fixation, water column denitrification, and zooplankton excretion are considered as well as the removal of NO3 by sedimentary denitrification. A global database of δ15NO3− observations is compiled from previous studies and compared to the model results on a regional basis where sufficient observations exist. The model is able to qualitatively and quantitatively reproduce many of the observed patterns such as high subsurface values in water column denitrification zones and the meridional and vertical gradients in the Southern Ocean. The observed pronounced subsurface minimum in the Atlantic is underestimated by the model presumably owing to too little simulated nitrogen fixation there. Sensitivity experiments reveal that algal NO3− uptake, nitrogen fixation, and water column denitrification have the strongest effects on the simulated distribution of nitrogen isotopes, whereas the effect from zooplankton excretion is weaker. Both water column and sedimentary denitrification also have important indirect effects on the nitrogen isotope distribution by reducing the fixed nitrogen inventory, which creates an ecological niche for nitrogen fixers and, thus, stimulates additional N2 fixation in the model. Important model deficiencies are identified, and strategies for future improvement and possibilities for model application are outlined.


1. Introduction

[2] Bioavailable nitrogen (fixed N) is one of the major limiting nutrients for algal photosynthesis, which drives the sequestration of CO2 from the surface ocean and atmosphere into the deep ocean via the sinking of organic matter. Changes in this so-called “biological pump” have been hypothesized to account for a significant amount of the glacial-interglacial fluctuations in atmospheric CO2 [McElroy, 1983; Falkowski, 1997]. However, the relative contributions of the biological and physical carbon pumps to CO2 variations remain controversial. The size of the oceanic fixed N inventory, which regulates the strength of the biological pump, is controlled by different biogeochemical processes that are difficult to constrain quantitatively in a global budget [Codispoti, 2007]. Nitrogen isotopes (both in dissolved and organic N species) in the water column and seafloor sediments are sensitive indicators of those processes [Brandes and Devol, 2002; Deutsch et al., 2004; Altabet, 2007].

[3] Many N transformational processes alter the ratio of the two stable forms of the nitrogen isotopes, 14N and 15N, differently, a process referred to as fractionation. Resulting variations in N isotopic composition can be described as deviations in 15N/14N ratio from an accepted standard

\[ \delta^{15}N = \left( \frac{15N/14N}{R_{std}} - 1 \right) \times 1000, \]  (1)

where \( R_{std} \) is the 15N/14N ratio of atmospheric N2 gas. Isotope fractionation can occur due to kinetic processes (i.e.,
different reaction rates for isotopes in a reactant product stream). It generally results in the enrichment of the heavier $^{15}$N isotope in the reaction substrate, and its depletion in the product. For example, preferential discrimination against $^{15}$N relative to $^{14}$N during algal NO$_3$ assimilation results in net enrichment of $^{15}$N in the residual NO$_3$ and net depletion of $^{15}$N in organic matter (OM). The degree of isotopic discrimination, or fractionation, for each process can be quantified with an enrichment factor, $\varepsilon = (^{15}k/^{14}k - 1) \times 1000$, where k is the specific reaction rate for each isotope [Mariotti et al., 1981].

[4] The predominant source and sink terms of the oceanic fixed N inventory, N$_2$ fixation and denitrification, respectively, have their own distinct effects on the signature of the N isotopes in the ocean. N$_2$ fixing prokaryotes (diazotrophs) introduce bioavailable N into the ocean close to that of atmospheric N$_2$ ($^{15}$N $\approx -2$–$0$‰) [Delwiche and Steyn, 1970; Minagawa and Wada, 1986; Macko et al., 1987; Carpenter et al., 1997]. Trichodesmium, one of the most important and best studied diazotrophs, blooms more frequently and extensively in warm ($>25^\circ$C) surface water where rates of aeolian Fe deposition are high such as the North Atlantic, Indian, and North Pacific compared to areas of low Fe deposition such as the South Pacific where the abundance of Trichodesmium appears to be much lower [Karl et al., 2002; Carpenter and Capone, 2008]. However, other unicellular diazotrophs have been observed to grow in cooler water near 20°C [Needoba et al., 2007], and it has been suggested that they also may significantly contribute to the global N$_2$ fixation rate [Zehr et al., 2001; Montoya et al., 2004].

[5] Denitrification occurs under suboxic conditions (O$_2$ < 5 $\mu$mol/kg) in the water column and in the seafloor sediments. Here, microbes use NO$_3$ instead of O$_2$ as the electron acceptor during respiration and convert it to gaseous forms of N (N$_2$O and N$_2$), which can then escape to the atmosphere [Codispoti and Richards, 1976]. The volume and distribution of suboxic water is affected by the temperature-dependent solubility of O$_2$ at the surface and the rate of subduction of oxygen-saturated water masses to greater depths, as well as the amount of organic matter that remineralizes in the ocean interior, both of which are sensitive to changes in climate. Anammox is another important process that occurs in anaerobic conditions and eliminates forms of fixed N (NO$_2$, NH$_3$) in the water column by converting them into N$_2$ gas [Mulder et al., 1995; Thamdrup and Dalsgaard, 2002; Kuyper et al., 2003]. It has been suggested that anammox may even eliminate more fixed N than water column denitrification in some oxygen minimum zones [Kuyper et al., 2005; Lam et al., 2009], but just how important of a role anammox plays in the global fixed N inventory has yet to be determined.

[6] Denitrifiers preferentially consume $^{14}$NO$_3$ leaving the residual oceanic NO$_3$ pool strongly enriched in the heavier $^{15}$N, with N isotope enrichment factors between 20–30‰ [Cline and Kaplan, 1975; Liu and Kaplan, 1989; Brandes et al., 1998; Altabet et al., 1999b; Voss et al., 2001]. Sedimentary denitrification is generally limited by the amount of NO$_3$ that diffuses into the reactive zones within the sediments. Therefore, it consumes nearly all of the influxing NO$_3$ available, leaving nearly unaltered $^{15}$N values in the overlying waters [Brandes and Devol, 1997, 2002; Sigman et al., 2003; Lehmann et al., 2004, 2007]. The average oceanic $^{15}$NO$_3$ value near 5‰ [Sigman et al., 1997, 1999] can be interpreted as the balance between the isotope effects of water column denitrification, sedimentary denitrification, and N$_2$ fixation [Brandes and Devol, 2002; Deutsch et al., 2004; Galbraith et al., 2004; Altabet, 2007].

[7] The $^{15}$N signal in the water column and seafloor sediments is also affected by fractionation processes within the food chain. Marine algae preferentially assimilate the lighter $^{14}$N into their biomass with a range of enrichment factors estimated in the field between 4–15‰ [Wada, 1980; Altabet et al., 1991, 1999b; Sigman et al., 1999; Altabet and Francois, 2001; Karsh et al., 2003; DiFiore et al., 2006]. Nitrogen is not lost or gained from the ocean during algal NO$_3$ assimilation, but the spatial separation between net assimilation and remineralization can cause a trend of decreasing $^15$NO$_3$ with depth. Distinguishing between the different isotope effects remains a challenge, especially in regions where multiple N transformational processes are occurring within close proximity.

[8] This study, for the first time to our knowledge, includes a dynamic nitrogen isotope module embedded within an existing global ocean-atmosphere-sea ice-biogeochemical model. This allows a direct comparison with nitrogen isotope observations, whereas previous box model studies could only be used more qualitatively [Giraud et al., 2000; Deutsch et al., 2004]. We provide a detailed description of the nitrogen isotope model and an assessment of its skill in reproducing present-day $^{15}$NO$_3$ observations. Comparison of model results with $^{15}$N observations will also be used to help to quantify processes that affect the global oceanic distribution of $^{15}$N. Sensitivity experiments illustrate the individual isotope effects of different processes on the spatial distribution of the N isotopes. In combination with measurements in ocean sediments and in the water column, the model can be a tool to better understand variations of $^{15}$N and the nitrogen cycle in the past and present.

2. Model Description

2.1. Physical Model

[9] The physical model is based on the University of Victoria Earth system climate model [Weaver et al., 2001], version 2.8. It includes a global, three-dimensional general circulation model of the ocean (Modular Ocean Model 2) with physical parameterizations such as diffusive mixing along and across isopycnals, eddy induced tracer advection [Gent and McWilliams, 1990] and a scheme for the computation of tidally induced diapycnal mixing over rough topography [Simmons et al., 2004]. Nineteen vertical levels are used with a horizontal resolution of 1.8° × 3.6°. To improve the simulation of equatorial currents, we have increased the meridional resolution in the tropics to 0.9° (between 10°S and 10°N and smoothly transitioning to 1.8° at 20°N/S) and added an anisotropic viscosity scheme [Large et al., 2001]. A more detailed description of this
parameterization and its effect on the equatorial circulation is provided in Text S1 of the auxiliary material. To account for the overestimated ventilation in the North Pacific, an artificial stratifying force equal to 0.04 Sv of freshwater is applied over the surface north of 55° in the Pacific and compensated elsewhere. A two dimensional, single level energy-moisture balance model of the atmosphere and a state-of-the-art dynamic-thermodynamic sea ice model are used, forced with prescribed NCEP/NCAR monthly climatological winds.

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**2.2. Marine Ecosystem Biogeochemical Model**

[10] The marine ecosystem model is an improved version of the 2N2PZD (2Nutrient, 2Phytoplankton, Zooplankton, Detritus) ecosystem model of [Schmittner et al., 2008] (Figure 1). The organic variables include two classes of phytoplankton, N2 fixing diazotrophs ($P_D$) and a “general” NO3 assimilating phytoplankton class ($P_O$), as well as zooplankton ($Z$) and organic detritus ($D$). The inorganic variables include dissolved oxygen ($O_2$) and two nutrients, nitrate ($NO_3^{-}$) and phosphate ($PO_4^{3-}$), both of which are consumed by phytoplankton and remineralized in fixed elemental ratios ($r_{N:P} = 16$, $r_{O:P} = 170$). We note, though,
that most diazotrophs have been found to have a $r_{N,P}$ of 50:1 and sometimes higher [e.g., Letelier and Karl, 1996, 1998; White et al., 2006]. This simplification is one of the reasons why the nitrogen surplus $N' = NO_3^- - 16PO_4^{3-}$ is generally underestimated in surface waters in the model (Figure S3). In addition to water column denitrification and $N_2$ fixation, we now include a parameterization for sedimentary denitrification (see auxiliary material equation (S11) and Figure 2), based on the flux of organic carbon into the seafloor sediments [Middelburg et al., 1996]. Since the model underestimates coastal upwelling, which drives large fluxes of organic carbon to the seafloor sediments, this parameterization is tuned to fit the global mean $\delta^{15}NO_3^-$ of 5‰ by multiplying the sedimentary denitrification equation by a constant factor ($a_{SD} = 4.5$). Global rates of model $N_2$ fixation, water column denitrification, and sedimentary denitrification are 101, 67.5, and 38.3 Tg N yr$^{-1}$, respectively. The relatively low model sedimentary to water column denitrification ratio of ~1:2 compared to other estimates from one-box models ranging from ~1:1 [Altabat, 2007] to ~4:1 [Brandes and Devol, 2002] is mostly due to the “dilution effect” [Deutsch et al., 2004], which reduces the “effective” fractionation effect of water column denitrification as $NO_3^-$ is locally consumed, an effect not incorporated in one-box models. This results in a lower sedimentary to water column denitrification ratio needed to set the global mean $\delta^{15}NO_3^-$ to 5‰ (see section 4.2 for further discussion). The complete marine ecosystem model description is provided in Text S2 of the auxiliary material. A comparison of the global distribution of $NO_3^-$, $O_2$, and $N'$ with World Ocean Atlas 2005 (WOA05) observations is shown in Figure S3.

11 Suboxic water, where water column denitrification occurs, is present in three main locations of the present-day oceans: the Eastern Tropical North Pacific (ETNP), the Eastern Tropical South Pacific (ETSP) and the Arabian Sea (Figure S3). Deficiencies in the physical circulation model simulate suboxic water in only one of these locations, the ETNP. The physical circulation model integrates coastal upwelling over a horizontal extent that is too large (due to its coarse resolution), which results in the underestimation of upwelling, export production, and the remineralization of organic matter at depth. This bias leads to too high $O_2$ concentrations, larger than required for water column denitrification, in the ETSP and the Arabian Sea. Suboxia in the so-called “shadow zone” of the ETNP is simulated better and investigated more in section 4.2. In the model, some water column denitrification also occurs in the Bay of Bengal and off SW Africa (Figure 2c), which has not been observed in the real ocean. However, the anammox reaction,
which also eliminates fixed N in the water column, has been found to occur off SW Africa [Knypers et al., 2005]. Naqvi [2008] measured low decomposition rates in the Bay of Bengal. Effective ballasting and scavenging of organic matter by the massive riverine input of terrestrial matter, an effect not included in the model, may prevent water column denitrification in the Bay of Bengal, which is close to suboxic.

[12] Diazotrophs grow according to the same principles as algal phytoplankton in the model (see Text S2), but we also account for some of their different characteristics. N$_2$ fixation breaks down the triple N bond of N$_2$, which is energetically more costly than assimilating fixed N [Holl and Montoya, 2005]. Therefore, in the model, the growth rate of diazotrophs is lower than that of general phytoplankton. Diazotrophs are not limited by NO$_3$ and will thrive in waters that are N deficient (i.e., low N′ as a result of denitrification) in which sufficient P and Fe are available. Denitrification and the propagation of N-deficient waters into the shallow thermocline by physical transport processes create an ecological niche for diazotrophs in the model, which stimulates N$_2$ fixation [Tyrrell, 1999].

[13] One of the most important and best studied diazotrophs, *Trichodesmium*, also has large iron (Fe) requirements for growth [Sanudo-Wilhelmy et al., 2001]. Diazotrophs may depend on aeolian Fe in oligotrophic waters because deep pycnocline inhibits upward mixing of subsurface Fe-replete waters into the euphotic zone [Falkowski, 1997; Karl et al., 2002]. Therefore, their growth rate is further reduced according to the Fe Limitation parameter (Figure 2a), where an estimate of aeolian dust deposition [Mahowald et al., 2005] is scaled between 0–1 by multiplying it by a constant factor and setting the maximum value to 1. This is a simple parameterization of Fe limitation of diazotrophy and its full effects are described elsewhere [Somes et al., 2010]. The majority of N$_2$ fixation in the model occurs in oligotrophic waters “downstream” of denitrification zones where sufficient Fe exists (i.e., via aeolian Fe deposition) (Figure 2b). The pattern of N$_2$ fixation (such as high values in the tropical/subtropical North Pacific, the western tropical/subtropical South Pacific, the western tropical/subtropical South Atlantic, the tropical/subtropical North Atlantic and the Indian Ocean) is mostly consistent with direct observations [e.g., Karl et al., 2002; Carpenter and Capone, 2008], with estimates based on the observed NO$_3$ deficit and simulated circulation [Deutsch et al., 2007], as well as with results from a more complex ecosystem model [Moore and Doney, 2007]. However, N$_2$ fixation in our model does not extend northward of 25–30°N in the North Pacific and North Atlantic, whereas some observations show N$_2$ fixation as far north as 35–40°N [Church et al., 2008; Kitajima et al., 2009].

### 2.3. Nitrogen Isotope Model

[14] The nitrogen isotope model simulates the distribution of the two stable nitrogen isotopes, $^{14}$N and $^{15}$N, in all N species throughout the global ocean that are included in the marine ecosystem model. Five prognostic variables of $\delta^{15}$N are embedded within the marine ecosystem model for all species containing nitrogen: NO$_3$, diazotrophs, algal phytoplankton, zooplankton and organic detritus (Figure 1). The ‘isotope effect’ is referred to in the following as the effect that each process has on the respective oceanic isotopic N pool, which depends on the $\delta^{15}$N value of the substrate, the process-specific enrichment factor ($\varepsilon$), and the degree of utilization ($u$) of the substrate during the reaction:

$$\delta^{15}\text{N}_{\text{product}} = \delta^{15}\text{N}_{\text{substrate}} - \varepsilon (1 - u)$$

where $u$ is the fraction of the initial substrate used in the reaction. For example, if all of the available substrate is consumed in the reaction (i.e., $u = 1$), the product will incorporate the $\delta^{15}$N value of the substrate, nullifying any potential fractionation. However, if the rate of utilization is low (i.e., $u < 1$), the product will incorporate a relatively light $\delta^{15}$N value compared to the substrate by the designated enrichment factor (Table 1).

[15] The processes in the model that fractionate nitrogen isotopes are algal NO$_3$ assimilation ($\varepsilon_{\text{ASSIM}} = 5\%$), zooplankton excretion ($\varepsilon_{\text{EXCR}} = 6\%$), and water column denitrification ($\varepsilon_{\text{WCD}} = 25\%$) (Table 1). Fractionation results in the isotopic enrichment of the more reactive, thermodynamically preferred, light $^{14}$N into the product of each reaction by a process-specific fractionation factor. For a detailed discussion of nitrogen isotope fractionation dynamics see Mariotti et al. [1981]. Although little fractionation occurs during N$_2$ fixation in the model, it has an important effect on $\delta^{15}$N by introducing relatively light atmospheric N$_2$ ($\delta^{15}$N = 0‰) into the oceanic fixed N inventory. Sedimentary denitrification also has been observed to have little effect on the oceanic isotopic N pool because denitrifiers consume nearly all NO$_3$ diffusing into the sediments [Brandes and Devol, 1997, 2002; Lehmann et al., 2004, 2007]. In the model, there is no fractionation during sedimentary denitrification ($\varepsilon_{\text{SD}} = 0\%$), although this is a simplification of observations [Lehmann et al., 2007]. Fractionation during the remineralization of organic matter is not included in the model. The complete nitrogen isotope model description is provided in Appendix A.

### 3. Nitrogen Isotope Model Results

[16] The model simulates complex spatial patterns of $\delta^{15}$NO$_3$ and $\delta^{15}$N organic matter (OM) throughout the global ocean (Figure 3, top). Patterns of surface $\delta^{15}$NO$_3$ and subsurface $\delta^{15}$N OM are similar but values are offset

<table>
<thead>
<tr>
<th>Process</th>
<th>Symbol</th>
<th>Model Enrichment Factor (%)</th>
<th>Field Estimates (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Algal NO$_3$ assimilation</td>
<td>$\varepsilon_{\text{ASSIM}}$</td>
<td>5</td>
<td>4–15</td>
</tr>
<tr>
<td>N$_2$ fixation</td>
<td>$\varepsilon_{\text{SFX}}$</td>
<td>1.5</td>
<td>0–2</td>
</tr>
<tr>
<td>Excretion</td>
<td>$\varepsilon_{\text{EXCR}}$</td>
<td>6</td>
<td>3–6</td>
</tr>
<tr>
<td>Water column denitrification</td>
<td>$\varepsilon_{\text{WCD}}$</td>
<td>25</td>
<td>22–30</td>
</tr>
<tr>
<td>Sedimentary denitrification</td>
<td>$\varepsilon_{\text{SD}}$</td>
<td>0</td>
<td>0–4</td>
</tr>
</tbody>
</table>

*aSee Appendix A for references.*
by two processes. First, as much as 5‰ offset due to fractionation during NO$_3^-$ uptake by phytoplankton and second, by fractionation during zooplankton excretion, which increases the δ$^{15}$N OM through zooplankton mortality (Figure 1). High δ$^{15}$NO$_3^-$ values (>15‰) are simulated in the eastern subtropical gyres, where surface NO$_3^-$ is depleted, and in regions in close proximity to simulated suboxic zones in the Eastern Pacific, Bay of Bengal, and Eastern Atlantic.

Figure 3. (top) Surface δ$^{15}$NO$_3^-$ and δ$^{15}$N of sinking detritus in the model. (bottom) Isotope effect sensitivity experiments where one isotope effect is neglected per simulation and its difference with CTL is shown to illustrate its individual effect on the CTL simulation.
(again, note that water column denitrification has not been actually observed in the Bay of Bengal and Eastern Atlantic). A clear interhemispheric asymmetry appears between the subtropical gyres of the Pacific and Atlantic with higher values of 14–20% simulated in the southern hemisphere and smaller values of 10–14% in the northern hemisphere. More intermediate $\delta^{15}$N values of 4–8% are found at high latitudes and near the equator where nutrient utilization is incomplete. $\delta^{14}$N minima (<4‰) are located in the western tropical/subtropical ocean basins, where N$_2$ fixation occurs in the model (Figure 2b). The remainder of this section presents a more quantitative description of the contributions of individual processes to these relatively complex spatial patterns of $\delta^{15}$NO$_3$ and $\delta^{15}$N OM.

[17] Figure 3 illustrates results from the full model (CTL) that includes all isotope effects (top panels) together with results from sensitivity experiments designed to isolate the effects of individual processes (bottom panels) on the global $\delta^{15}$N distribution. This is accomplished by removing the isotope effect of one process per experiment and then calculating the difference ($\Delta \delta^{15}$N) with CTL. In the “NO$_3$ Assimilation” and “Excretion” experiments, the enrichment factors $\varepsilon_{\text{ASSIM}}$ and $\varepsilon_{\text{EXCR}}$, respectively, are set to zero. In the “$N_2$ fixation” experiment the diazotroph’s N isotope ratio is set equal to that of other phytoplankton at each location. In the “Water Column Denitrification” and “Sedimentary Denitrification” experiments, the entire process is switched off (thereby changing the global N inventory). These latter experiments also show the indirect effect that both denitrification processes have on $\delta^{15}$N through the stimulation of $N_2$ fixation. In all other isotope effect experiments, the total N inventory does not change.

### 3.1. Algal NO$_3$ Assimilation

[18] As phytoplankton preferentially assimilate $^{14}$NO$_3$ into organic matter, the residual inorganic N pool becomes enriched in $^{15}$NO$_3$. This creates an offset between surface $\delta^{15}$NO$_3$ and $\delta^{15}$N OM that sinks toward the seafloor, which is set by the enrichment factor for NO$_3$ assimilation ($\varepsilon_{\text{ASSIM}} = 5\%$) (“NO$_3$ Assimilation” experiment, Figure 3). The surface NO$_3$ utilization effect is also affected by the extent to which NO$_3$ is depleted. When NO$_3$ utilization is low (i.e., NO$_3$‐replete water exists), which occurs in High Nitrate Low Chlorophyll (HNLC) regions such as the Southern Ocean, the subarctic North Pacific, and the eastern equatorial Pacific, surface $\delta^{15}$NO$_3$ is determined by the source of $\delta^{15}$NO$_3$ being supplied to the surface. Algae will fractionate this NO$_3$ during assimilation near the full extent set by the designated enrichment factor because of the abundance of available NO$_3$. In this oceanographic setting, the expected $5\%$ difference between $\delta^{15}$NO$_3$ and $\delta^{15}$N OM is almost fully expressed (i.e., $\delta^{15}$N‐PO$_4 = \delta^{15}$NO$_3 – \varepsilon_{\text{ASSIM}}$ with $\mu_{\text{ASSIM}} \approx 0$ in equation (2)). Thus, surface NO$_3$ utilization in HNLK regions has a small influence on the surface $\delta^{15}$NO$_3$ signature, but play an important role for $\delta^{15}$N OM that sinks out of the euphotic zone. This is perhaps most obvious in the Southern Ocean and in the subarctic North Pacific where $\Delta \delta^{15}$NO$_3$ is small, whereas $\Delta \delta^{15}$N OM is strongly negative (“NO$_3$ Assimilation” experiment, Figure 3).

[19] A different response is observed in oligotrophic regions where surface NO$_3$ is depleted. Once the algae consume nearly all available NO$_3$ (which itself becomes enriched in $^{15}$N), they acquire the same N isotope signature from the source NO$_3$ (i.e., $\delta^{15}$N‐PO$_4 = \delta^{15}$NO$_3$ with $\mu_{\text{ASSIM}}$ approaching 1). This drives the high $\delta^{15}$N values in both NO$_3$ and OM in the subtropics with maxima in the eastern poleward edges of the gyres (Figure 3). Although $\delta^{15}$NO$_3$ values are very high there, they have a small effect on $\delta^{15}$N elsewhere because NO$_3$ concentrations are very low. For instance, when low NO$_3$ water mixes with nearby water with significantly higher NO$_3$, the resulting $\delta^{15}$NO$_3$ value will be weighted toward the water parcel containing more NO$_3$ [see also Deutsch et al., 2004]. This ‘dilution effect’ prevents high $\delta^{15}$NO$_3$ values in regions with high surface NO$_3$ utilization from having a large impact on the $\delta^{15}$NO$_3$ signature across the nitracline.

### 3.2. Denitrification

[20] Denitrification only occurs at depth but its isotope effect can reach the surface due to upwelling and vertical mixing. Water column denitrification has a large enrichment factor and displays a very strong N isotope effect in close proximity to the simulated suboxic zones in the Eastern Pacific, Bay of Bengal, and Eastern Atlantic (“Water Column Denitrification” experiment in Figure 3). The unresolved poleward undercurrents along the western continental margin of the Americas (which could, in the real world, propagate high $\delta^{15}$NO$_3$ away from the sub-surface suboxic zones [Kienast et al., 2002]) may restrict the simulated water column denitrification isotope effect too much to regions proximal to the suboxic zones. Both water column and sedimentary denitrification also indirectly lead to lower $\delta^{15}$NO$_3$ values “downstream” of denitrification zones because they create N‐deficient water that stimulates additional $N_2$ fixation, which introduces low $\delta^{15}$N into the ocean. This negative feedback also decreases the horizontal extension of high $\delta^{15}$NO$_3$ signature originating from suboxic zones, because $N_2$ fixation introduces much lower $\delta^{15}$N into the ocean.

### 3.3. $N_2$ Fixation

[21] The addition of newly fixed, isotopically light atmospheric $N_2$ ($\delta^{15}N_2 = 0$) by diazotrophs is the reason for the low $\delta^{15}$N values in the western tropical/subtropical ocean basins. Since denitrification is the only process in the model that creates N‐deficient water, and therefore an ecological niche for diazotrophs, the majority of N$_2$ fixation in the model occurs “downstream” of denitrification zones after phytoplankton have consumed all remaining surface NO$_3$ and where sufficient aeolian Fe deposition exists. This low $\delta^{15}$NO$_3$ signature is evident in the subtropical North/South Pacific, the subtropical North/South Atlantic, and the Bay of Bengal (“$N_2$ Fixation” experiment, Figure 3).

### 3.4. Excretion

[22] According to our model results, the N isotope effect of excretion has a smaller influence on the simulated distribution of $\delta^{15}$N in the global ocean (“Excretion” experiment, Figure 3) compared to the other processes discussed.
Seasonal sampling biases exist depending on the region. More details on the data sets can be found in the respective ocean region subsections that follow. Comparisons are presented for the Southern Ocean (Indian-Pacific sector), the Eastern Tropical North Pacific, and the Subtropical North Atlantic. Other regions with available $^{15}\text{N}$ observations included in the data set but not discussed in the text are the Bering Sea [Lehmann et al., 2005], the Northeast Pacific [Galbraith, 2006], the Arabian Sea [Altabet et al., 1999a], the Central Equatorial Pacific [Somes et al., 2010], and the eastern Pacific sector of the Southern Ocean [Sigman et al., 1999].

[24] Global measures of model performance for $^{15}\text{N}$ are presented in Table 2. These measures should be interpreted taken into account the highly localized nature of some of the processes as well as the limited regions covered by the database. A displacement in the location of denitrification, for example, will lead to a large decrease in the correlation coefficient and a large increase in the RMS errors. The CTL model has a correlation coefficient of 0.68, implying that the model explains 46% of the variance in the observations. The decrease of the correlation coefficient and the increase of the RMS error due to the neglect of a particular process can be regarded as the importance that this process plays in explaining the global $^{15}\text{N}$ observations of the database. The correlation coefficient measures the pattern of variability and neglects the absolute values, whereas the RMS error considers the deviation of the model from the observations in absolute values. Neglecting water column denitrification leads to the largest decrease in the correlation coefficient to 0.29 and to the second largest increase in the RMS error after N$_2$ fixation. Neglecting N$_2$ fixation and algal NO$_3^-$ assimilation lead to the next largest decrease in the correlation coefficient.

[25] Sedimentary denitrification or excretion is not included, then the correlation coefficients decrease similarly, with both having relatively weaker effects on the distribution of $^{15}\text{NO}_3^-$. Then, according to these measures, water column denitrification is the most important process determining the global $^{15}\text{NO}_3^-$ distribution of available observations in the database, followed by N$_2$ fixation and algal NO$_3^-$ assimilation, respectively. Finally, sedimentary denitrification and excretion are the least important.

### 4.1 Southern Indian-Pacific Ocean

The Southern Ocean represents a critical region of biogeochemical cycling in the ocean because it is the largest open ocean region with incomplete drawdown of the major nutrients. This results in an excess amount of CO$_2$ at the

**Table 2. Global Measures of $^{15}\text{NO}_3^-$ Model Performance**

<table>
<thead>
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<th>Model</th>
<th>$r$</th>
<th>$P$</th>
<th>RMS</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>0.68</td>
<td>&lt;0.0001</td>
<td>0.73</td>
</tr>
<tr>
<td>Algal NO$_3$ assimilation</td>
<td>0.60</td>
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<tr>
<td>N$_2$ fixation</td>
<td>0.52</td>
<td>0.0026</td>
<td>2.1</td>
</tr>
<tr>
<td>Excretion</td>
<td>0.65</td>
<td>0.00010</td>
<td>0.80</td>
</tr>
<tr>
<td>Water column denitrification</td>
<td>0.29</td>
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<tr>
<td>Sedimentary denitrification</td>
<td>0.64</td>
<td>0.00010</td>
<td>0.82</td>
</tr>
</tbody>
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*aCorrelation coefficient ($r$), correlation significance ($P$), and root mean squared (RMS) error normalized by the standard deviation of the observations.*
surface, which is released to the atmosphere (under preindustrial conditions). The degree to which surface nutrients are utilized here may have profound impacts on ocean-atmosphere exchanges of CO2. Figure 5 shows a comparison with observations recorded in the region [Sigman et al., 1999; Altabet and Francois, 2001; DiFiore et al., 2006]. This data subset compiles observations from 8 cruises covering various seasons. Since all cruises do not cover the same location, some seasonal biases can be expected, yet, we still decided to use annual averages for maximum spatial coverage. Note the model does not simulate interannual variability due to the prescribed monthly climatological winds and temporally constant biogeochemical parameters.

[26] Qualitatively, the inverse trend of increasing $\delta^{15}$NO$_3^-$ with decreasing NO$_3^-$ (Figure 5a) is reproduced by the model. However, the slope is underestimated suggesting that the enrichment factor for algal NO$_3^-$ assimilation used in the model (\(\varepsilon_{\text{ASSIM}} = 5\%\)) is too low, in agreement with DiFiore et al. [2006] that suggests at least 7\%. The simulated vertical gradient is in good agreement with the observations. Deep water $\delta^{15}$NO$_3^-$ at 2000 m depth is around 5\% and slowly increasing throughout the lower pycnocline to around 6\% at 500 m depth. The model slightly underestimates $\delta^{15}$NO$_3^-$ between 100 m depth by $\sim$0.2\%, whereas near surface values are underestimated by 1.5\%.

[27] The meridional gradient of observed surface $\delta^{15}$NO$_3^-$, with low values of $\sim$5\% at high latitudes and higher values of $\sim$10\% north of 45\°S, are captured by the model, except that the gradient in the model is sharper and shifted northward by about 10°. Very high $\delta^{15}$NO$_3^-$ values are simulated north of $\sim$40°S off the southern coast of Australia (Figure 5c). This is due to the fact that the model overestimates the utilization of surface NO$_3^-$ relative to observations there (Figures 5c and S3). Where the simulated NO$_3^-$ is almost completely consumed (i.e., NO$_3^-$ < 1 \muM) (see Figure 5c contour line), the remaining $\delta^{15}$NO$_3^-$ values become as high as 18\%. Since none of the existing $\delta^{15}$NO$_3^-$ observations was collected in such low NO$_3^-$ concentrations (Figure 5a), it is impossible, at this time, to falsify this aspect of the N isotope model response. We note this heavy $\delta^{15}$NO$_3^-$ signature in these low NO$_3^-$ waters has little effect on $\delta^{15}$N across the nitracline in the model because the $\delta^{15}$N signature of very low NO$_3^-$ water becomes diluted out as it mixes with much higher NO$_3^-$ water (see section 3.1).

4.2. Eastern Tropical North Pacific

[28] The ETNP contains the largest suboxic zone in the ocean, where water column denitrification occurs. The relatively small spatial scale of suboxic zones makes them difficult for the model to simulate accurately and deficiencies in the coarse resolution physical model prevent it from fully resolving some important physical processes, especially in coastal regions. Underestimating coastal upwelling (due to coarse resolution) results in corresponding underestimation of primary production, organic matter remineralization, and O$_2$ consumption at depth. This is a major...
reason for overestimated dissolved O$_2$ at depth in areas with significant coastal upwelling (e.g., off Peru and NW Mexico) (Figure S3), too large for water column denitrification to occur. Preliminary experiments suggest that increased vertical resolution can improve the simulation of productivity and suboxia in the Eastern Tropical South Pacific (not shown).

[29] The ability to reproduce the equatorial undercurrents that transport relatively oxygen-rich water from the western basin is also important for the simulation of the Eastern Pacific suboxic zones. The anisotropic viscosity scheme [Large et al., 2001] improves equatorial dynamics considerably (Text S2 and Figure S1). The Pacific Equatorial Undercurrent increases from 0.15 m/s to nearly 0.8 m/s, just slightly weaker than observations, which show velocities near 1 m/s (Figure S2). The North Equatorial Countercurrent in the model also displays lower current velocities than observed, and does not deliver enough oxygen-rich water directly to the ETNP suboxic zone. This is likely the main reason why the model requires a relatively low sedimentary to water column denitrification ratio of 1:2 to set the global mean $\delta^{15}$NO$_3$ compared to estimates from one-box models [Brandes and Devol, 2002; Altabet, 2007], which cannot account for any important effects that occur locally within the denitrification zone. However, note that our model significantly overestimates NO$_3$ consumption via water column denitrification in the ETNP compared to observations (Figure 6a). Therefore, it is likely that our sedimentary to water column denitrification ratio of 1:2 is too low, but it does highlight the importance that the NO$_3$ consumption/dilution effect can have on determining the global mean $\delta^{15}$NO$_3$.

[31] Figure 6 shows model $\delta^{15}$NO$_3$ compared to observational $\delta^{15}$NO$_3$ data collected during November 1999 [Sigman et al., 2005] and October 2000 (M. Altabet, unpublished data, 2010). The model captures the general observed trend of increasing $\delta^{15}$NO$_3$ as NO$_3$ is consumed during water column denitrification (Figure 6a). The model’s too low N:P ratio for diazotrophs ($r = 16:1$ where...
observations show $r = \sim 50:1$ [White et al., 2006; Letelier and Karl, 1996, 1998]) may partly explain its incapacity to simulate some of the relatively high N:P values of observations. The range of simulated values is also likely to be more limited compared to the observations due to the missing interannual and synoptic climate variability in the model. Figure 6b compares the horizontally averaged $\delta^{15}$NO$_3$ depth profiles only within the hypoxic zone (O$_2 < 10$ μM) at 300 m (contoured on Figure 6c) to account for the displaced OMZ. Within this region, the model is able to capture the general vertical distribution of $\delta^{15}$NO$_3$ seen in the measured data, such as the surface minimum and subsurface maximum. [32] $\delta^{15}$NO$_3$ in the ETNP decreases toward the surface [Cline and Kaplan, 1975; Brandes et al., 1998; Voss et al., 2001; Sigman et al., 2005] suggesting a source of isotopically light N at the surface. Brandes et al. [1998] proposed that in the Arabian Sea as much as 30% of primary production must be supported by N$_2$ fixation in order to account for the low surface $\delta^{15}$NO$_3$. Other observations also suggest that the decrease in $\delta^{15}$NO$_3$ toward the surface is likely due to the fixation of atmospheric N$_2$ and the subsequent, closely coupled remineralization-nitrification cycle [Sigman et al., 2005]. We test this hypothesis by comparing the observations with the model experiment in which the isotope effect of N$_2$ fixation is neglected (“NO NFIX”). In this case, the model overestimates surface $\delta^{15}$NO$_3$ by $\sim 12\%$ (Figure 6b) and the surface minimum is not simulated. This experiment demonstrates that the input of isotopically light fixed N from N$_2$ fixation in the model best explains the decreasing trend of $\delta^{15}$NO$_3$ observations toward the surface. In the model, 20% of the fixed N loss via denitrification is reintroduced into the surface by N$_2$ fixation occurring directly above the denitrification zone in the ETNP. The fact that the difference between the subsurface maximum and the near surface minimum is underestimated in the model (6% versus 8% in the observations) suggests that in the real world the locally reintroduced fraction could be larger than 20%.

4.3. North Atlantic

[33] Uncertainties regarding processes that can affect the nitrogen isotope signal make it challenging to interpret and simulate nitrogen isotopes in the North Atlantic. Estimates of atmospheric N deposition [Duce et al., 2008] and the assimilation-mineralization-nitrification cycle are not well constrained. Although atmospheric N deposition may be significant in this region [Michaels et al., 1996; Lipschultz et al., 2002; Knapp et al., 2005, 2008], its isotopic composition is not well known and therefore is not included in the model at this time. Figure 7 shows the comparison of annual model $\delta^{15}$NO$_3$ with available observations from cruises in May 2001 and 2004 (M. Altabet and J. P. Montoya, unpublished data, 2010), October 2002 [Knapp et al., 2008], and May 2005 [Bourbonnais et al., 2009]. The model overestimates the $\delta^{15}$NO$_3$ values everywhere, by 0.9‰ on average and by 2‰ at 200 m depth, presumably due mostly to the underestimation of N$_2$ fixation, but possibly also because atmospheric N deposition and/or fractionation during the remineralization of organic matter are not included. Both of these processes would act to decrease subsurface values of $\delta^{15}$NO$_3$. Underestimated N’ in the North Atlantic (Figure S3) also indicates too little N$_2$ fixation, but we again note the too low N:P ratio for diazotrophs also contributes to this N’ underestimation to some degree.

[34] N$_2$ fixation is most likely underestimated in the model because it does not consider dynamic elemental cycling of the microbial loop. It has been suggested that DOP is more labile relative to DON and recycles through the microbial loop more efficiently, which can help relieve diazotrophs of P limitation in this region and enhance N$_2$ fixation [Wu et al., 2000]. The model is able to reproduce the pattern of low $\delta^{15}$NO$_3$ in the thermocline qualitatively, just not quantitatively to the extent present in the observations. Sedimentary denitrification in the North Atlantic sti-
mulates enough N2 fixation in the model to generate a subsurface δ15NO3− minimum. When sedimentary denitrification is switched off (‘No SD’), the thermocline minimum is weaker and agrees less with the observations. This suggests that sedimentary denitrification is an important factor influencing N2 fixation in the Subtropical North Atlantic, but not the only factor.

5. Discussion and Conclusions

[35] A new model of nitrogen isotopes has been implemented into the three-dimensional ocean component of a global Earth system climate model capable of millennial timescale simulations. Despite some model deficiencies, we have shown that this model can successfully reproduce the general spatial patterns of δ15NO3− measured in the ocean. Sensitivity experiments allowed us to isolate the individual N isotope effects of various N transformational processes on the global distribution of δ15N. Algal NO3− assimilation, water column denitrification, and N2 fixation all have strong influences in setting the global patterns of δ15NO3− in the ocean, whereas the effect of zooplankton excretion is weaker.

[36] These simulations show that the isotope effect of algal NO3− assimilation can drive very large spatial gradients in both δ15NO3− and δ15N OM depending on the ocean environment (Figure 3). In HNLC areas where surface NO3− utilization is low and algae are able to fractionate NO3− at their designated enrichment factor, the δ15N OM signature decreases. However, when NO3− utilization is high, the δ15N OM signature is more similar to the δ15NO3− value it consumes because the effective degree of fractionation becomes much lower (see section 3.1). Surface NO3− utilization gradients can transition rapidly, for example due to changes in ocean circulation or atmospheric Fe deposition, and can possibly drive large and rapid changes in δ15NO3− and δ15N OM. The important influence of surface NO3− utilization on the global distribution of N isotopes in the model suggests that changes in surface NO3− utilization patterns throughout Earth’s history could contribute to large fluctuations in δ15N observed in sediment records, especially near fronts where large surface NO3− gradients exist [see also Altabet and Francois, 1994; Farrell et al., 1995; Sigman et al., 1999; Brunelle et al., 2007; Galbraith et al., 2008; Robinson and Sigman, 2008].

[37] The model simulates a strong direct and indirect isotope effect of denitrification. High δ15NO3− produced by water column denitrification has clear regional impacts and is also responsible for overall elevated δ15NO3− of the ocean relative to the N2 fixation source (see below). The indirect effect of both water column and sediment denitrification is mediated by the production of N-deficient water, which creates an ecological niche for diazotrophs. This stimulates additional N2 fixation when other suitable conditions for N2 fixation also exist (e.g., warm (>20°C), N-depleted water with sufficient P and Fe). This indirect effect also attenuates the horizontal circulation of high δ15NO3− waters, originating from regions of water column denitrification, which causes its direct isotope effect to be regionalized near suboxic zones in the model.

[38] Key features of the model have been identified that are in need of further development. The coarse resolution physical circulation model does not fully resolve the dynamics of coastal upwelling regimes, which in part drive the flux of organic matter toward the seafloor sediments and its remineralization in the water column, as well as indirectly influences ventilation of suboxic zones. This is critical in the simulation of water column denitrification and sedimentary denitrification, which are important processes with respect to the global N isotope balance. Future model versions will include additional vertical levels to better resolve continental shelves as well as higher horizontal resolution. The model neglects dynamic elemental stoichiometry such as high N:P ratios of diazotrophs and the more efficient recycling of DOP relative to DON in microbial loops, which can help relieve diazotrophs of their P limitation and allow them to fix additional N2 into the oceanic fixed N pool in oligotrophic waters. The ecosystem model also suffers from the exclusion of Fe as a prognostic tracer preventing it from being able to simulate differences in ecosystems limited by marconutrients (NO3−, PO4−3) versus micronutrients (Fe).

[39] Future applications of this model will include simulations of past climates, and direct comparison with δ15N sediment records will be used to test the model results. This approach may be useful to quantify past interactions between the marine N cycle and its isotopes, as well as their impact on climate, and may provide new insights into important physical and biogeochemical changes throughout Earth’s history.

Appendix A: Nitrogen Isotope Model

[40] The open system fractionation equation is used for fractionation during algal NO3− assimilation [Altabet and Francois, 2001]:

\[
\delta^{15}N-P_{O} = \delta^{15}NO_{3}^{-} - \varepsilon_{ASSIM}(1 - \nu_{NO3}),
\]

where δ15N-P_O is the δ15N of phytoplankton biomass assimilated during one time step, Δt, and ϵASSIM is the fraction of available NO3− that is converted into biomass (uASSIM = J_P/PO4 × Δt/NO3). When algae assimilate all available NO3− into their biomass (i.e., uASSIM = 1) they will incorporate the same δ15N value as that of the source material. Many studies have estimated the fractionation factor in both laboratory and ocean environments. A wide variety of values have been reported in culture settings ranging from 0.7‰ to 23‰ [Wada and Hattori, 1978; Montoya and McCarthy, 1995; Waser et al., 1998; Needoba et al., 2003; Granger et al., 2004]. A more confined range has been observed in field estimates from 4‰ to 15‰ [Wada, 1980; Altabet et al., 1991, 1999b; Sigman et al., 1999; Altabet and Francois, 2001; Karsh et al., 2003; DiFiore et al., 2006]. In our model we choose a constant value of 5‰ which is near the majority of estimates, although it is important to bear in mind the uncertainty in the parameter choice and the possibility that it varies in space and time.

[41] Nitrate in suboxic waters have been observed to have much higher δ15N values due to fractionation during denitrification. Observations from present-day suboxic zones in
the Eastern Tropical North Pacific (ETNP) and the Arabian Sea (AS) have reported fractionation factors ranging from 22 to 30‰ [Cline and Kaplan, 1975; Liu and Kaplan, 1989; Brandes et al., 1998; Altabet et al., 1999b; Voss et al., 2001]; we adopt a value of 25‰ in the model. Note that because these estimates were derived from field studies in which the isotope effect was estimated from the total nitrogen loss, they implicitly include the effect of anammox [Galbraith et al., 2008]. Fractionation during denitrification is also simulated using the open system fractionation equation

$$\delta^{15}NO_3 = \delta^{15}NO_3^\text{ox} - \varepsilon_{WCD}(1 - u_{NO3}),$$  \hspace{1cm} (A2)

where NO$_3^{15}$X is the oxygen-equivalent reduction of nitrate converted into N$_2$ gas during denitrification. The term $u_{DENI}$ is the fraction of available NO$_3$ which is reduced into N$_2$ gas ($u_{NO3} = \mu_D \times 0.8 \times r_{NO_3} \times \rho_{NO3}^{\text{sox}} \times L_{NO3} \times \Delta t/\Delta NO_3$).

[42] Excretion is the process responsible for the stepwise enrichment of $^{15}$N along the trophic chain in our model and is simulated using the instantaneous fractionation equation:

$$\delta^{15}NO_3 = \delta^{15}Z - \varepsilon_{EXCR}.$$  \hspace{1cm} (A3)

The instantaneous fractionation equation is used because excretion will always be a small fraction of the total zoo plankton biomass and has been measured to be depleted by $\sim 6$‰ relative to its body [Montoya, 2008], which is the source of the excreted nitrogen. This leads to the average enrichment of $\sim 3.4$ per trophic level [Minagawa and Wada, 1984].

[43] Implementing these fractionation equations into the marine ecosystem model requires us to consider the exchanges of $^{14}$N and $^{15}$N between the various N pools separately. Total nitrogen abundance now has the form

$$N = ^{14}N + ^{15}N$$  \hspace{1cm} (A4)

for each variable in the isotope model. A fractionation coefficient is calculated for each process so the same equations for total N can be applied to $^{15}$N [Giraud et al., 2000]. For example, consider fractionation during algal NO$_3^-$ assimilation. The isotopic ratio of new nitrogen biomass ($P_D$) is found using equations (1) and (2):

$$\delta^{15}N-P_D = \beta_{ASSIM} \delta^{15}N-P_O$$  \hspace{1cm} (A5)

where

$$\beta_{ASSIM} = \frac{^{15}NO_3}{^{14}NO_3} - \frac{\varepsilon_{ASSIM}(1 - u_{NO3})R_{nd}}{1000}.\hspace{1cm} (A6)$$

[44] Applying equations (A4) and (A5) gives the amount of new $\delta^{15}N-P_O$ relative to the amount of total new nitrogen biomass, which is given by the primary production ($J_O P_O$), calculated by the marine ecosystem model:

$$\delta^{15}N-P_O = \frac{\beta_{ASSIM}}{1 + \beta_{ASSIM}} J_O \delta^{15}N-P_O.$$  \hspace{1cm} (A7)

Analogous derivations can be done for all fractionation coefficients. The time-dependent set of equations for $^{15}$N which are embedded into the marine ecosystem model are as follows:

$$\frac{\partial^{15}NO_3}{\partial t} = \left( R_{D_{H2}O} + \frac{\beta_{EXCR}}{1 + \beta_{EXCR}} \gamma Z + R_{P_{H2}P_O} - \frac{\beta_{ASSIM}}{1 + \beta_{ASSIM}} u_{NO3} J_O P_D \right) \times \left[ 1 - \frac{\beta_{WCD}}{1 + \beta_{WCD}} 0.8 R_{N/P_{sox}}^{NO3} L_{NO3} \right].$$  \hspace{1cm} (A8)

$$\frac{\partial^{15}P_O}{\partial t} = \frac{\beta_{ASSIM}}{1 + \beta_{ASSIM}} J_O P_D - R_{P_{H2}P_O} - R_{P_{H2}GP_O} Z - R_{P_{H2}P_{sox}} P_O^2,$$  \hspace{1cm} (A9)

$$\frac{\partial^{15}N-P_D}{\partial t} = \left( \frac{\beta_{ASSIM}}{1 + \beta_{ASSIM}} u_{N} + \frac{\beta_{NFIX}}{1 + \beta_{NFIX}} (1 - u_{N}) \right) J_O P_D$$

$$- R_{P_{H2}GP_D} Z - R_{P_{H2}P_{sox}} P_D.$$  \hspace{1cm} (A10)

$$\frac{\partial^{15}N-Z}{\partial t} = \gamma_1 \left[ R_{P_{H2}GP_O} + R_{P_{H2}GP_D} Z \right] - \frac{\beta_{EXCR}}{1 + \beta_{EXCR}} \gamma Z - R_{Z_{H2}} Z^2,$$  \hspace{1cm} (A11)

$$\frac{\partial^{15}N-D}{\partial t} = \left( 1 - \gamma_1 \right) \left[ R_{P_{H2}GP_O} + R_{P_{H2}GP_D} Z + R_{P_{H2}P_{sox}} P_D \right]$$

$$+ R_{P_{H2}P_{sox}} P_D^2 + R_{Z_{H2}} Z^2 - R_{D_{H2}O} D - R_{D_{wD}} D^2.$$  \hspace{1cm} (A12)

where $R_{X - P_{O}, P_{D}, Z_{D}} = ^{15}N/(^{14}N + ^{15}N)$ is the ratio of heavy over total nitrogen. The complete parameter description is provided in Text S2. Here it suffices to note that the equations for total nitrogen ($^{14}N + ^{15}N$) are identical to the ones of $^{15}$N except that $R_X = \beta_X/(1 + \beta_X) = 1$ in the total nitrogen equations.

[45] The model was carefully tested with zero fractionation in order to quantify and minimize numerical errors, which can occur for example due to slightly negative values of biological tracers caused by inaccuracies of the advection scheme. The biological code was adjusted to avoid negative concentrations as much as possible. Initial numerical errors in $\delta^{15}$N ranged from $\pm 1$‰ in grid points at the sea floor to $\pm 0.1$‰ in the upper ocean. Setting $R_{nd} = 1$ instead of $R_{nd} = 0.0036765$, the actual atmospheric N$_2$ isotope ratio, reduces the numerical errors by over an order of magnitude. This prevents $^{15}$N from becoming very close to zero as often, where inaccuracies of the advection scheme can cause it to be negative. This modification amounts to a scaling of $^{15}$N and $^{14}$N by a constant factor which does not affect the $\delta^{15}$N dynamics. The remaining numerical errors of $\pm 0.1$‰ in the deep ocean and $\pm 0.01$‰ in the upper ocean are
2 orders of magnitude smaller than the observed variability. The model was integrated for over 5,000 years as it approaches equilibrium.

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